

A Research on Chloroplast Gene Transformation

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Article Received: 11 May 2019

Article Accepted: 17 August 2019

Article Published: 18 October 2019

ABSTRACT

Up to a number of useful genes transformed into plants via nuclear transformation. But the nuclear transformation of crop plants has some perspective problems those over chloroplast transformation. Many useful genes have been transformed into a chloroplast genome. Tobacco plastid has been transformed for the production of high levels of vaccines, antigens, biopharmaceuticals etc. The chloroplast is a useful platform for expression of recombinant protein in higher plants. Transgenes can be introduced into the plastid genome (either by PEG of plant protoplasts or commonly by biolistic method). The chloroplast of all eukaryotic transformation algae and embryophytes (bryophytes, ferns and seed plants) are the product of a singular endosymbiosis event that happened approximately 1.5 billion years ago and occurred through the uptake of a cyanobacterium by a heterotrophic protist.

Abbreviations: (PEG) transformation - poly ethylene glycol transformation, (GFP) - Green fluorescent protein.

Keywords: Chloroplast, Gene Silencing, biopharmaceuticals, Biolistic transformation, protoplast, PEG, Agrobacterium.

INTRODUCTION

The chloroplast is also called plastids in plant cells and eukaryotes algae (Wani et al. 2010; Wang, Yin, and Hu 2009). About 1.5 billion years ago, chloroplast came from endosymbiosis during that time when cyanobacterial cell was engulfed heterotrophic eukaryote (Adem, Beyene, and Feyissa 2017a; Bock, n.d.; Maliga 2003). plastid transgene is high and recombinant protein denote up to 70% of leaf protein (Adem et al., 2017a; Day & Goldschmidt-Clermont, 2011; Oey et al., 2009; Ruhlman, Verma, Samson, & Daniell, 2010). A nucleus is bidirectional which is called anterograde regulation. Nucleus coded regulator transfer information of cell types expresses proteins that are the main factor for plastids function within particular cell types (Jung and Chory 2010; Leister 2005; Cullis et al. 2009; Haque et al. 2018). Three types of genomes are set up in plant cells in some plants, two of this genome are transformable nucleus genome and genome of chloroplasts. The genome of the chloroplast is energetically photosynthetically seeded plants those have genome 120-220Kb and coding gene about 120-130kb (Bock 2014; Oey et al. 2009).

Chloroplast act as metabolic midpoints in cellular reactions to signals and respond to retrograde signaling. The process changes solar energy to carbohydrate by the process of photosynthesis and releases CO₂ during such process. Photosynthesis is the central function of plastids. They play important roles in plants physiology and improvement, containing vitamins, photohormones, fatty acid and an overabundance of metabolites and gathering of nitrogen and sulfur (Daniell et al. 2016; Bobik and Burch-smith 2015; Arlen et al. 2008). Chloroplast acts important role in number of critical metabolic process such as photosynthesis, fatty acid, amino acid and production of unimportant metabolites. chloroplast also like as plastid derived from endosymbiosis of inherited cyanobacterium in an eukaryotic cell and cause gene conversion between nucleus and chloroplast (Bansal and Saha 2012; Gould, Waller, and Mcfadden 2008; Green 2011; Maliga, Bock, and Brunswick 2011; Reyes-prieto, Weber, and Bhattacharya 2007). About 100 chloroplasts in one leaf every cell have up to 100 plastome copies and such multi-ploidy gives high levels of foreign genes (C. Cui, Song, Tan, Zhou, Zhao, Ma, Liu, Hussain, Wang, et al. 2011; Cosa et al. 2015; Allen 2015). Current evidence indicates that chloroplast has nearly 2,000 and 3,000 most

important localized proteins that are encoded by host genome (Barry et al. 2012; Armbruster et al. 2011; Klee 2010a). Presentation of plastid transformation benefit in production of drug or biofuels (Day and Goldschmidt-Clermont 2011; Manuscript 2012; Gould, Waller, and Mcfadden 2008; Daniell et al. 2010; Specht, Miyake-Stoner, and Mayfield 2010; Wang, Yin, and Hu 2009; Verma and Daniell 2007; Protoc et al. 2008). For engineering of agronomic traits and bioformaceuticals, translastomic technology is useful in plant and metabolic engineering (Singh, Verma, and Bansal 2010).

The chloroplast is key important organelle of plants and green algae, harbors their genome is blamable many important purposes like photosynthesis, lipid metabolism, starch and amino acid biosynthesis (Armbruster et al., 2011). Excessive quantity assembly of recombinant protein in chloroplast has a deleterious effect on plant growth (Gottschalk et al. 2016; Liu and Peng 2013). Additional oxidative control is third machinery which produce oxidative oxygen species (ROS), hydrogen peroxide (H₂O₂) and singlet oxygen (O-1) that cause specific changes in nuclear expression (Estavillo et al. 2013; Jajic, Sarna, and Strzalka 2015; Kopcowski 2013; Suzuki et al. 2012; Miller, Shulaev, and Mittler 2008). very central studies of plastid genome –encoded genes and ORFS have been approved by genetic method preliminary with production of a transformation vector in which a selectable marker removed the marker gene in plant plastome (Scotti et al. 2011; Allison, Simon, and Maligal 1996; Thiele et al. 2008).

Different plastids have different definite characteristic and role such as, proplast (existing in meristematic region of the plants); chloroplasts (existing in petal of florae and ovaries which are colored plastids capable to collect great quantity of carotenoids); amyloplasts (existent in storage tissues such as roots and seed endosperm); etioplasts (exist in cotyledon of dusky green angiosperm seedlings); elaioplasts (lipid Storing plastids); leucoplast (less pigment -plastids exist in root cells); chloroplast (chlorophyll comprising plastids helpful in photosynthesis (Rogalski et al. 2015; Bock, n.d.; Egea et al. 2010; Olejniczak et al. 2016). chloroplast transformation commanded 410-fold rise the expression of the same protein.

In alteration of chloroplast next measures take place: light observation and consequent expression of nuclear and plastids genes, biosynthesis of lipid and pigments, introduction of proper pigment-binding proteins such as chlorophyll a/b-binding proteins into evolving chloroplasts, addition of these proteins into the thylakoid membranes, and protein association into efficient multiplexes (Rudowska et al. 2012; López-Juez 2007; Waters and Langdale 2009). Chloroplast DNA moves from cell to cell to tissue grafts freshly determine (Gutiérrez et al. 2012; Waters and Langdale 2009).

Transformation Methods

An ordinary method used in *Agrobacterium* transformation frequently prime random insertion of transgenes. Later addition of imported genes, chloroplast transformation have been useful apparatus helpful of the plastid coded proteins through the formation of knockout and site path mutant (Arlen et al. 2008). Two types of RNA-polymerases are required for the transcription of coded-proteins, both of which essential for biogenesis (Tiller and Bock 2014). *Chlamydomonas* chloroplast transformation succeeded by mixing cell wall-deficient cell with the DNA interesting presence glass-drops (Potvin and Zhang 2010; Daniell, Khan, and Allison 2002; Olejniczak et al.

2016). before few decade Of wheat chloroplast transformation has been withdrawn (Hanson et al. 2012; C. Cui, Song, Tan, Zhou, Zhao, Ma, Liu, Hussain, and Wang 2011) . In 1990, chloroplast transformation was obtained in tobacco(Scotti et al. 2011) .

In green microalgae, *Chlamydomonas reinhardtii* earliest time chloroplast transformation took place(Y. Cui, Qin, and Jiang 2014; Blowers et al. 1989; Day and Goldschmidt-Clermont 2011). The major marker used for chloroplast transformation is known as the *aadd* gene(Y. Cui, Qin, and Jiang 2014; Goldschmidt-clermont 1991; Svab and Maliga 1993). Many methods are tangled in chloroplast transformation are species-specific or heterologous chloroplast transformation vector established in a way that margins the foreign gene and inserts them through homologous recombination at a determined and accurate position in plastome(Adem, Beyene, and Feyissa 2017a). For *P.subcordiformis* chloroplast transformation two plastids pPSC-GFP and pPSCB were hand-me-down(Doron, Segal, and Shapira 2016).

Alga preparation and bombardment factors were as before defines. Golds particle about 0.8 -1.5 mm in diameters were used as plastid carriers, for every bombardment 2-3 mg plasmid DNA was used. The imported gene to be inserted into plastid genome is edged by left and right nucleotide sequences from hot plastid genome which determines the of transgene delivered by homologous recombination(Klee 2010a; Wang, Yin, and Hu 2009). Nearly 16 location of plastid genome were used for definitely directing of transgene addition in plastid genomes(Klee 2010b; Preview 2017). Two approaches for the preparation of chloroplast vectors have been established lately (1) Gateway system (2) Vector design(Jin and Daniell 2015; Vafae et al. 2014).

Biolistic and polyethylene glycol (PEG) mediated protoplast transformation was use nearly 20 years back(Bock 2014; Maliga, Bock, and Brunswick 2011).chloroplast vector used To attain effective chloroplast transformation, the constant integration of Transgene is by homologous recombination of adjoining sequence used in vectors (chloroplast vectors)(Beeramganti, Beeramganti, and Subramanyam 2012). Cabbage has regeneration and transformation efficiently totally depend on the genotypes (Beeramganti, Beeramganti, and Subramanyam 2012; Culture 2015). Another policy was created based on the Cre-loxP site-specific recombination system by Kuroda and Maliaga (Scotti et al. 2011; Fleischmann et al. 2011). The transgenic plant has been forthcoming like " green bio factories " for the collection of recombinant products(Gutiérrez et al. 2012; Rajan and Howard 2012). Engineer complex metabolic pathways have the opportunity to transfer or mass multiple genes(Gutiérrez et al. 2012; Cosa et al. 2015).

IMPORTANT ADVANTAGES OF CHLOROPLAST TRANSFORMATION

Collection of foreign protein

Chloroplast transformation efficient to collect a greater quantity of foreign protein(Verma and Daniell 2007).

High level of transgene expression

Chloroplast transformation upturn high level of transgene expression with the reduction in epigenetic inactivation of transgene and cause of parental inheritance (Saha and Fibres 2014; Bock 2001; Maliga, Bock, and Brunswick 2011; Cosa et al. 2015).

Creation and cultivation of genetically modified plants

Use of chloroplast is beneficial then chloroplast DNA is not conveyed through pollens, which stops gene drive from the heritably improved plant to other plants. This is an important device for the formation and farming of genetically modified plants (Beeramganti, Beeramganti, and Subramanyam 2012).

Gene silencing and position effect

Chloroplast transformation prevents gene silencing and position effect. (Y. Cui et al., 2014; Garg et al., 2002).

An important role in the production of pharmaceuticals and vaccine:

Chloroplast transformation plays important roles in the production of pharmaceuticals and particular subunit vaccine (Gutiérrez et al. 2012; Olejniczak et al. 2016; Maliga, Bock, and Brunswick 2011; Lo 2011; Adem, Beyene, and Feyissa 2017b).

Co-expression multiple gene systems

Ability to co-expression multiple genes in operons which is necessary for those vaccines that involve multiple epitopes for their suitable purpose (Lo 2011; Y. Cui, Qin, and Jiang 2014).

Chloroplast transformation is helpful in molecular farming, metabolic engineering, engineering resistance, experimental evolution and in vivo analysis of gene expression (Bock, n.d.).

APPLICATIONS OF CHLOROPLAST TRANSFORMATION

Vaccine production and biopharmaceuticals

More than 90% of the global population cannot manage to pay for insulin, a drug is required to treat global diabetes dominant (Mall 2007). The drugs made by the plant chloroplast overcome more tasks as they do not need expensive manufacture and can be stored without losing effectiveness (Holtz et al. 2015).

The hyper appearance of vaccine antigens or therapeutic proteins in the transgenic chloroplast, chloroplast or antibiotic-free selection systems available in plastid transformation system became successful in the oral delivery of vaccine antigens against anthrax, cholera, tetanus and canine parvovirus (Maliga 2003). Various therapeutic proteins have been produced from a single plant *Chlamydomonas reinhardtii* (Almaraz-delgado et al. 2014).

Biomaterials and industrial enzyme production

Biomaterials and important enzymes have been positively formed by the chloroplast genome engineering. Metabolic engineering using the chloroplast genome produced the highest level of the poly (P-hydro benzoic acid) polymer (25% dry weight in normal healthy plants (Snell 2011).

The production of renewable recyclable plastic poly hydroxyl butyrate by using an optimized genetic concept for the plastid transformation of tobacco was designed using an operon extension policy (Beilen and Poirier 2008). Different biomaterials and enzymes have been engineered via the chloroplast genome of tobacco (Guda and Daniell 2000).

Production of biofuels

Effective enzymes are essential for lignocellulosic biofuel production. Due to high-cost levels of enzymes, technical issues and lack of infrastructure, the production of ethanol, derived from cellulose is limited. So, chloroplast transformation is helpful in biofuel production. (Jin and Daniell 2015). Plants are used as bio factories to produce appropriate recombinant enzymes in an alternative to microbial fermentation (Pantaleoni et al. 2014).

CONCLUSION

Chloroplast genome has become the goal of many plant genetic transformation efforts due to its huge advantages over nuclear genome of plant. Addressing the above barriers will make chloroplast genome very attractive site for various biotechnological application with unbelievable impact on human life. For introduction of agronomic traits and biopharmaceutical plants, plastid transformation technology has gained peak importance. Next future plastid transformation technique will be supplied for a wide range of purposes both in basic and applied. Recent studies brings to light the importance of generating plastid transformation systems using somatic embryos or embryogenic calli, which might pave the way to engineer the plastid genome of several major crops in which regeneration is mediated through somatic embryogenesis. Due to high level of gene expression, chloroplast genome transformation is an emerging biological tool for medical importance.

REFERENCES

- Adem, Muhamed, Dereje Beyene, and Tileye Feyissa. 2017a. "Recent Achievements Obtained by Chloroplast Transformation." *Plant Methods* 13 (1): 1–11. <https://doi.org/10.1186/s13007-017-0179-1>.
- "Recent Achievements Obtained by Chloroplast Transformation." *Plant Methods* 13 (1): 1–11. <https://doi.org/10.1186/s13007-017-0179-1>.
- Allen, John F. 2015. "Why Chloroplasts and Mitochondria Retain Their Own Genomes and Genetic Systems : Colocation for Redox Regulation of Gene Expression." <https://doi.org/10.1073/pnas.1500012112>.
- Allison, Lori A, Lee D Simon, and Pal Maligal. 1996. "Deletion of RpoB Reveals a Second Distinct Transcription System in Plastids of Higher Plants" 15 (11): 2802–9.
- Almaraz-delgado, Alma Lorena, José Flores-uribe, Víctor Hugo Pérez-españa, and Edgar Salgado-manjarrez. 2014. "Production of Therapeutic Proteins in the Chloroplast of *Chlamydomonas Reinhardtii*," 1–9.
- Arlen, Philip A., Michael Singleton, Jeffrey J. Adamovicz, Yi Ding, Abdolreza Davoodi-Semiromi, and Henry Daniell. 2008. "Effective Plague Vaccination via Oral Delivery of Plant Cells Expressing F1-V Antigens in Chloroplasts." *Infection and Immunity* 76 (8): 3640–50. <https://doi.org/10.1128/IAI.00050-08>.
- Armbruster, Ute, Paolo Pesaresi, Mathias Pribil, Alexander Hertle, and Dario Leister. 2011. "Update on Chloroplast Research: New Tools , New Topics , and New Trends." *Molecular Plant* 4 (1): 1–16. <https://doi.org/10.1093/mp/ssq060>.
- Bansal, Kailash C., and Dipnarayan Saha. 2012. "Chloroplast Genomics and Genetic Engineering for Crop Improvement." *Agricultural Research* 1 (1): 53–66. <https://doi.org/10.1007/s40003-011-0010-6>.

- Barry, C. S., G. M. Aldridge, G. Herzog, Q. Ma, R. P. McQuinn, J. Hirschberg, and J. J. Giovannoni. 2012. "Altered Chloroplast Development and Delayed Fruit Ripening Caused by Mutations in a Zinc Metalloprotease at the Lutescent2 Locus of Tomato." *Plant Physiology* 159 (3): 1086–98. <https://doi.org/10.1104/pp.112.197483>.
- Beeramganti, Navaneeth, Haritha Beeramganti, and Koona Subramanyam. 2012. "Chloroplast Expression Vector System & Its Transformation" 2 (4): 47–58.
- Beilen, Jan B Van, and Yves Poirier. 2008. "Production of Renewable Polymers from Crop Plants," 684–701. <https://doi.org/10.1111/j.1365-313X.2008.03431.x>.
- Blowers, Alan D, Lawrence Bogorad, Katherine B Shark, and John C Sanford. 1989. "Studies on Chlamydomonas Chloroplast Transformation : Foreign DNA Can Be Stably Maintained in the Chromosome" 1 (January): 123–32.
- Bobik, Krzysztof, and Tessa M Burch-smith. 2015. "Chloroplast Signaling within , between and beyond Cells" 6 (October): 1–26. <https://doi.org/10.3389/fpls.2015.00781>.
- Bock, Ralph. n.d. "Advances in Chloroplast Transformation and Biotechnological Applications The Plastid (Chloroplast) Genome." "Engineering Plastid Genomes : Methods , Tools , and Applications in Basic Research and Biotechnology." <https://doi.org/10.1146/annurev-arplant-050213-040212>.
- "Transgenic Plastids in Basic Research and Plant Biotechnology." <https://doi.org/10.1006/jmbi.2001.4960>.
- "Genetic Engineering of the Chloroplast: Novel Tools and New Applications." *Current Opinion in Biotechnology* 26: 7–13. <https://doi.org/10.1016/j.copbio.2013.06.004>.
- Cosa, Brandy De, William Moar, Seung-bum Lee, Michael Miller, Henry Daniell, Electron Microscopy, and Imaging Facility. 2015. "HHS Public Access" 19 (1): 71–74. <https://doi.org/10.1038/83559>. Overexpression.
- Cui, Cuiju, Fei Song, Yi Tan, Xuan Zhou, Wen Zhao, Fengyun Ma, Yunyi Liu, Javeed Hussain, Yuesheng Wang, et al. 2011. "Stable Chloroplast Transformation of Immature Scutella and Inflorescences in Wheat (*Triticum Aestivum* L.)." *Acta Biochimica et Biophysica Sinica* 43 (4): 284–91. <https://doi.org/10.1093/abbs/gmr008>.
- "Stable Chloroplast Transformation of Immature Scutella and Inflorescences in Wheat (*Triticum Aestivum* L .)," no. February: 284–91. <https://doi.org/10.1093/abbs/gmr008>. Advance.
- Cui, Yulin, Song Qin, and Peng Jiang. 2014. "Chloroplast Transformation of *Platymonas* (*Tetraselmis*) *Subcordiformis* with the Bar Gene as Selectable Marker." *PLoS ONE* 9 (6): 1–7. <https://doi.org/10.1371/journal.pone.0098607>.
- Cullis, C. A., B. J. Vorster, C. Van Der Vyver, and K. J. Kunert. 2009. "Transfer of Genetic Material between the Chloroplast and Nucleus: How Is It Related to Stress in Plants?" *Annals of Botany* 103 (4): 625–33. <https://doi.org/10.1093/aob/mcn173>.
- Culture, Plant Tissue. 2015. "In Vitro Regeneration of Eight Cultivars of *Brassica Oleracea* Var . *Capitata*," 80–87. <https://doi.org/10.1007/s11627-014-9648-7>.

Daniell, Henry, Muhammad S. Khan, and Lori Allison. 2002. "Milestones in Chloroplast Genetic Engineering: An Environmentally Friendly Era in Biotechnology." *Trends in Plant Science* 7 (2): 84–91. [https://doi.org/10.1016/S1360-1385\(01\)02193-8](https://doi.org/10.1016/S1360-1385(01)02193-8).

Daniell, Henry, Choun Sea Lin, Ming Yu, and Wan Jung Chang. 2016. "Chloroplast Genomes: Diversity, Evolution, and Applications in Genetic Engineering." *Genome Biology* 17 (1): 1–29. <https://doi.org/10.1186/s13059-016-1004-2>.

Daniell, Henry, Nameirakpam D Singh, Hugh Mason, and Stephen J Streatfield. 2010. "NIH Public Access" 14 (12): 669–79. <https://doi.org/10.1016/j.tplants.2009.09.009.Plant-made>.

Day, Anil, and Michel Goldschmidt-clermont. 2011. "The Chloroplast Transformation Toolbox : Selectable Markers and Marker Removal," 540–53. <https://doi.org/10.1111/j.1467-7652.2011.00604.x>.

Day, Anil, and Michel Goldschmidt-Clermont. 2011. "The Chloroplast Transformation Toolbox: Selectable Markers and Marker Removal." *Plant Biotechnology Journal* 9 (5): 540–53. <https://doi.org/10.1111/j.1467-7652.2011.00604.x>.

Doron, Lior, Na Segal, and Michal Shapira. 2016. "Transgene Expression in Microalgae — From Tools to Applications Technical Approaches Used For" 7 (April): 1–24. <https://doi.org/10.3389/fpls.2016.00505>.

Egea, Isabel, Cristina Barsan, Wanping Bian, Eduardo Purgatto, Alain Latché, Christian Chervin, Mondher Bouzayen, and Jean-claude Pech. 2010. "Chromoplast Differentiation : Current Status and Perspectives Mini Review" 51 (10): 1601–11. <https://doi.org/10.1093/pcp/pcq136>.

Estavillo, Gonzalo M., Kai Xun Chan, Su Yin Phua, and Barry J. Pogson. 2013. "Reconsidering the Nature and Mode of Action of Metabolite Retrograde Signals from the Chloroplast." *Frontiers in Plant Science* 3 (January): 1–9. <https://doi.org/10.3389/fpls.2012.00300>.

Fleischmann, Tobias T, Lars B Scharff, Sibah Alkatib, Sebastian Hasdorf, and Mark A Scho. 2011. "Nonessential Plastid-Encoded Ribosomal Proteins in Tobacco : A Developmental Role for Plastid Translation and Implications for Reductive Genome Evolution" 23 (September): 3137–55. <https://doi.org/10.1105/tpc.111.088906>.

Garg, A. K., J.-K. Kim, T. G. Owens, A. P. Ranwala, Y. D. Choi, L. V. Kochian, and R. J. Wu. 2002. "Trehalose Accumulation in Rice Plants Confers High Tolerance Levels to Different Abiotic Stresses." *Proceedings of the National Academy of Sciences* 99 (25): 15898–903. <https://doi.org/10.1073/pnas.252637799>.

Goldschmidt-clermont, Michel. 1991. "Transgenic Expression of Aminoglycoside Adenine Transferase in the Chloroplast : A Selectable Marker for Site- Directed Transformation of Chlamydomonas" 19 (15): 4083–89.

Gottschamel, Johanna, Andreas Lössl, Stephanie Ruf, Yanliang Wang, Morten Skaugen, Ralph Bock, and Jihong Liu Clarke. 2016. "Production of Dengue Virus Envelope Protein Domain III-Based Antigens in Tobacco Chloroplasts Using Inducible and Constitutive Expression Systems." *Plant Molecular Biology* 91 (4–5): 497–512. <https://doi.org/10.1007/s11103-016-0484-5>.

- Gould, Sven B, Ross F Waller, and Geoffrey I Mcfadden. 2008. "Plastid Evolution," no. January. <https://doi.org/10.1146/annurev.arplant.59.032607.092915>.
- Green, Beverley R. 2011. "Chloroplast Genomes of Photosynthetic Eukaryotes." *Plant Journal* 66 (1): 34–44. <https://doi.org/10.1111/j.1365-313X.2011.04541.x>.
- Guda, C, and H Daniell. 2000. "Stable Expression of a Biodegradable Protein-Based Polymer in Tobacco Chloroplasts," 257–62.
- Gutiérrez, Carla L., Javier Gimpel, Carolina Escobar, Sergio H. Marshall, and Vitalia Henríquez. 2012. "Chloroplast Genetic Tool for the Green Microalgae *Haematococcus Pluvialis* (Chlorophyceae, Volvocales)." *Journal of Phycology* 48 (4): 976–83. <https://doi.org/10.1111/j.1529-8817.2012.01178.x>.
- Hanson, Maureen R, Benjamin N Gray, Beth A Ahner, and Adriana Chiappetta. 2012. "Chloroplast for Engineering of Photosynthesis In *Posidonia Transformation Oceanica* Cadmium Induces Changes in DNA Methylation and Chromatin Patterning" 63 (2). <https://doi.org/10.1093/jxb/err313>.
- Haque, Effi, Hiroaki Taniguchi, Md. Mahmudul Hassan, Pankaj Bhowmik, M. Rezaul Karim, Magdalena Śmiech, Kaijun Zhao, Mahfuzur Rahman, and Tofazzal Islam. 2018. "Application of CRISPR/Cas9 Genome Editing Technology for the Improvement of Crops Cultivated in Tropical Climates: Recent Progress, Prospects, and Challenges." *Frontiers in Plant Science* 9 (May): 1–12. <https://doi.org/10.3389/fpls.2018.00617>.
- Holtz, Barry R, Brian R Berquist, Lindsay D Bennett, Vally J M Kommineni, Ranjith K Munigunti, Earl L White, Don C Wilkerson, Kah-yat I Wong, Lan H Ly, and Sylvain Marcel. 2015. "Commercial-Scale Biotherapeutics Manufacturing Facility for Plant-Made Pharmaceuticals," 1180–90. <https://doi.org/10.1111/pbi.12469>.
- Jajic, Ivan, Tadeusz Sarna, and Kazimierz Strzalka. 2015. "Senescence, Stress, and Reactive Oxygen Species," 393–411. <https://doi.org/10.3390/plants4030393>.
- Jin, Shuangxia, and Henry Daniell. 2015. "The Engineered Chloroplast Genome Just Got Smarter." *Trends in Plant Science* 20 (10): 622–40. <https://doi.org/10.1016/j.tplants.2015.07.004>.
- Jung, Hou-sung, and Joanne Chory. 2010. "Signaling between Chloroplasts and the Nucleus : Can a Systems Biology Approach Bring Clarity to a Complex and Highly Regulated Pathway ?" 152 (February): 453–59. <https://doi.org/10.1104/pp.109.149070>.
- Klee, Harry J. 2010a. "Improving the Flavor of Fresh F." *New Phytologist* 187: 44–56. <https://doi.org/10.1111/j.1469-8137.2010.03281.x>.
- "Tansley Review Improving the Flavor of Fresh Fruits : Genomics , Biochemistry , And," 44–56.
- Kopczewski, Tomasz. 2013. "Redox Signals as a Language of Interorganellar Communication in Plant Cells" 8 (12): 1153–63. <https://doi.org/10.2478/s11535-013-0243-4>.
- Leister, Dario. 2005. "Genomics-Based Dissection of the Cross-Talk of Chloroplasts with the Nucleus and Mitochondria in *Arabidopsis*" 354: 110–16. <https://doi.org/10.1016/j.gene.2005.03.039>.

- Liu, Jihong, and Clarke Peng. 2013. "Plant Biotechnology for Food Security and Bioeconomy," 1–3. <https://doi.org/10.1007/s11103-013-0097-1>.
- Lo, Andreas G. 2011. "Chloroplast-Derived Vaccines against Human Diseases : Achievements , Challenges and Scopes," 527–39. <https://doi.org/10.1111/j.1467-7652.2011.00615.x>.
- López-Juez, Enrique. 2007. "Plastid Biogenesis, between Light and Shadows." *Journal of Experimental Botany* 58 (1): 11–26. <https://doi.org/10.1093/jxb/erl196>.
- Maliga, Pal. 2003. "Progress towards Commercialization of Plastid Transformation Technology" 21 (1): 20–28.
- Maliga, Pal, Ralph Bock, and New Brunswick. 2011. "Plastid Biotechnology : Food , Fuel , and Medicine for The" 155 (April): 1501–10. <https://doi.org/10.1104/pp.110.170969>.
- Mall, R Andall L S. 2007. "Comparison of whole chloroplast genome sequences to choose noncoding regions for phylogenetic studies in angiosperms: the tortoise and the hare III 1" 94 (3): 275–88. <https://doi.org/10.3732/ajb.94.3.275>.
- "NIH Public Access" 23 (5): 238–45. <https://doi.org/10.1016/j.tibtech.2005.03.008.Breakthrough>.
- Miller, Gad, Vladimir Shulaev, and Ron Mittler. 2008. "Reactive Oxygen Signaling and Abiotic Stress," 481–89. <https://doi.org/10.1111/j.1399-3054.2008.01090.x>.
- Oey, Melanie, Marc Lohse, Bernd Kreikemeyer, Ralph Bock, Am Mu, and Hospital Hygiene. 2009. "Exhaustion of the Chloroplast Protein Synthesis Capacity by Massive Expression of a Highly Stable Protein Antibiotic," 436–45. <https://doi.org/10.1111/j.1365-313X.2008.03702.x>.
- Olejniczak, Szymon Adam, Ewelina Łojewska, Tomasz Kowalczyk, and Tomasz Sakowicz. 2016. "Chloroplasts: State of Research and Practical Applications of Plastome Sequencing." *Planta* 244 (3): 517–27. <https://doi.org/10.1007/s00425-016-2551-1>.
- Pantaleoni, Laura, Paolo Longoni, Lorenzo Ferroni, Costanza Baldisserotto, and Sadhu Leelavathi. 2014. "Chloroplast Molecular Farming : Efficient Production of a Thermostable Xylanase by Nicotiana Tabacum Plants and Long-Term Conservation of the Recombinant Enzyme," 639–48. <https://doi.org/10.1007/s00709-013-0564-1>.
- Potvin, Gabriel, and Zisheng Zhang. 2010. "Strategies for High-Level Recombinant Protein Expression in Transgenic Microalgae: A Review." *Biotechnology Advances* 28 (6): 910–18. <https://doi.org/10.1016/j.biotechadv.2010.08.006>.
- Preview, *Plant Physiology*. 2017. "RESEARCH REPORT Short Title: Plastid Transformation In" 0029. <https://doi.org/10.1104/pp.17.00857>.
- Protoc, Nat, Dheeraj Verma, Nalapalli P Samson, Vijay Koya, and Henry Daniell. 2008. "A Protocol for Expression of Foreign Genes in Chloroplasts," no. June 2014: 739–58. <https://doi.org/10.1038/nprot.2007.522>.

Rajan, Vidya, and John A Howard. 2012. "Overproduction of Recombinant Proteins in Plants Overproduction of Recombinant Proteins in Plants Erin Egelkrout , Vidya Rajan , John A . Howard *," no. March 2016. <https://doi.org/10.1016/j.plantsci.2011.12.005>.

Reyes-prieto, Adrian, Andreas P M Weber, and Debashish Bhattacharya. 2007. "The Origin and Establishment of the Plastid in Algae and Plants," no. June: 147–68. <https://doi.org/10.1146/annurev.genet.41.110306.130134>.

Rogalski, Marcelo, Leila do Nascimento Vieira, Hugo P. Fraga, and Miguel P. Guerra. 2015. "Plastid Genomics in Horticultural Species: Importance and Applications for Plant Population Genetics, Evolution, and Biotechnology." *Frontiers in Plant Science* 6 (July): 1–17. <https://doi.org/10.3389/fpls.2015.00586>.

Rudowska, Łucja, Katarzyna Gieczewska, Radosław Mazur, Maciej Garstka, and Agnieszka Mostowska. 2012. "Chloroplast Biogenesis - Correlation between Structure and Function." *Biochimica et Biophysica Acta - Bioenergetics* 1817 (8): 1380–87. <https://doi.org/10.1016/j.bbabi.2012.03.013>.

Ruhlman, T., D. Verma, N. Samson, and H. Daniell. 2010. "The Role of Heterologous Chloroplast Sequence Elements in Transgene Integration and Expression." *Plant Physiology* 152 (4): 2088–2104. <https://doi.org/10.1104/pp.109.152017>.

Saha, Dipnarayan, and Allied Fibres. 2014. "Chloroplast Genomics and Genetic Engineering for Crop Improvement Chloroplast Genomics and Genetic Engineering for Crop Improvement," no. March 2012. <https://doi.org/10.1007/s40003-011-0010-6>.

Scotti, Nunzia, Daniela Gargano, Paolo Lenzi, and Teodoro Cardi. 2011. "Transformation of the Plastid Genome in Higher Plants CHAPTER 7 Transformation of the Plastid Genome in Higher Plants," no. January.

Singh, A. K., S. S. Verma, and K. C. Bansal. 2010. "Plastid Transformation in Eggplant (*Solanum Melongena* L.)." *Transgenic Research* 19 (1): 113–19. <https://doi.org/10.1007/s11248-009-9290-z>.

Snell, Kristi D. 2011. "Corresponding Author :." <https://doi.org/10.1104/pp.110.169581>.

Specht, Elizabeth, Shigeki Miyake-Stoner, and Stephen Mayfield. 2010. "Micro-Algae Come of Age as a Platform for Recombinant Protein Production." *Biotechnology Letters* 32 (10): 1373–83. <https://doi.org/10.1007/s10529-010-0326-5>.

Suzuki, Nobuhiro, Shai Koussevitzky, R O N Mittler, and G A D Miller. 2012. "ROS and Redox Signalling in the Response of Plants to Abiotic Stress," 259–70. <https://doi.org/10.1111/j.1365-3040.2011.02336.x>.

Svab, Zora, and P A L Maliga. 1993. "High-Frequency Plastid Transformation in Tobacco" 90 (February): 913–17.

Thiele, Wolfram, Waltraud X Schulze, Ralph Bock, Marcelo Rogalski, and Mark A Scho. 2008. "Rpl33 , a Nonessential Plastid-Encoded Ribosomal Protein in Tobacco , Is Required under Cold Stress Conditions" 20 (20): 2221–37. <https://doi.org/10.1105/tpc.108.060392>.

Tiller, Nadine, and Ralph Bock. 2014. "The Translational Apparatus of Plastids and Its Role in Plant Development." *Molecular Plant* 7 (7): 1105–20. <https://doi.org/10.1093/mp/ssu022>.

Vafae, Yavar, Agata Staniek, Maria Mancheno-solano, and Heribert Warzecha. 2014. "A Modular Cloning Toolbox for the Generation of Chloroplast Transformation Vectors" 9 (10). <https://doi.org/10.1371/journal.pone.0110222>.

Verma, D., and H. Daniell. 2007. "Chloroplast Vector Systems for Biotechnology Applications." *Plant Physiology* 145 (4): 1129–43. <https://doi.org/10.1104/pp.107.106690>.

Wang, Huan Huan, Wei Bo Yin, and Zan Min Hu. 2009. "Advances in Chloroplast Engineering." *Journal of Genetics and Genomics* 36 (7): 387–98. [https://doi.org/10.1016/S1673-8527\(08\)60128-9](https://doi.org/10.1016/S1673-8527(08)60128-9).

Wani, Shabir H, Nadia Haider, Hitesh Kumar, and N B Singh. 2010. "Plant Plastid Engineering," 500–512.

Waters, Mark T., and Jane A. Langdale. 2009. "The Making of a Chloroplast." *EMBO Journal* 28 (19): 2861–73. <https://doi.org/10.1038/emboj.2009.264>.